

绵羊 *FecB* 基因编码蛋白的 序列、结构和功能分析

Sequence, structure and function analysis of the protein
encoded by the Sheep *FecB* gene

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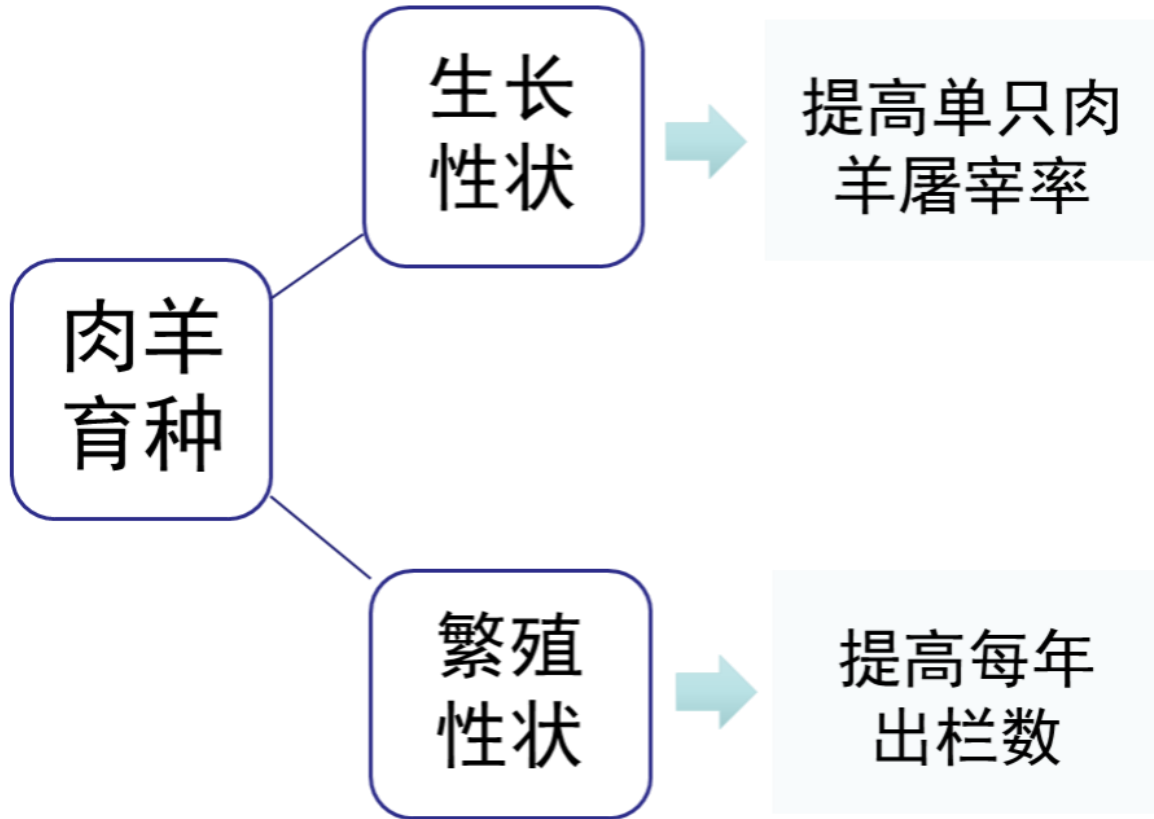
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研究展望

一. 研究背景：绵羊多羔的意义



绵羊的繁殖性状是绵羊的**重要经济性状**，为数量性状，属低遗传力性状。它包括受胎率、多胎性（产羔率及每胎产羔数）及羔羊存活率3个方面，其中**多胎性**是决定养羊业效益的最主要的性状之一，也是绵羊业多产高产的基础。

一. 研究背景：绵羊多羔主效基因 *FecB* 的发现

1919 年，澳大利亚 B.Sear 在新南威尔士州中毛型美利奴羊中选出一只高繁殖力母羊，并把它的后代作为一个分离群，他的侄儿 J. Sear 和 D.Sear 兄弟继续对高繁殖力性状进行选择，并以其牧场的名字命名为 Booroola 美利奴羊。

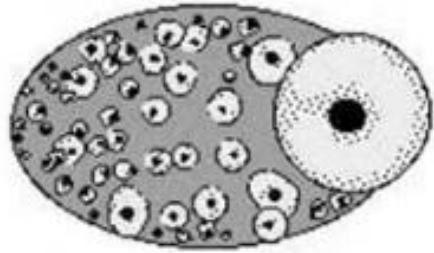
1958 年，澳大利亚联邦科学与工业研究组织用 Seears 兄弟 Booroola 羊群 1 只公羊和 13 只一胎产三羔和一胎产四羔的母羊继续扩繁而成为今天的 Booroola 绵羊。

1980 年，澳大利亚和新西兰进行的研究证实了 Booroola 绵羊高繁殖力属单基因遗传，通常认为这一遗传突变是由点突变、重复或缺失引起的，之后将该主效基因命名为 F。

1989 年，该基因被国际绵羊和山羊遗传命名委员会命名为 *FecB* (既 Fec=fecundity, B=Booroola)

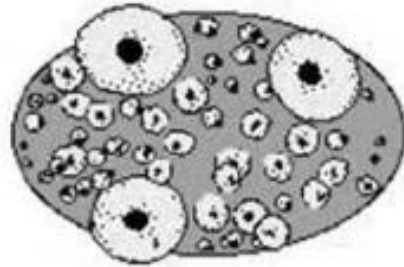


一. 研究背景: *FecB*对表型影响



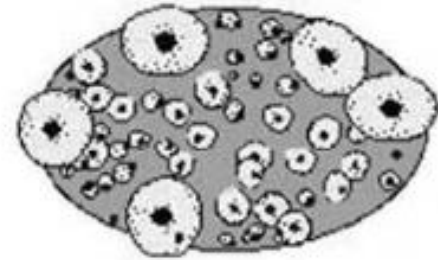
Wild type +/+

Ovulation rate = 1
Size of ovulatory follicles 7mm



Heterozygous +/B

Ovulation rate = 3
Size of ovulatory follicles 4.5mm



Homozygous B/B

Ovulation rate = 5
Size of ovulatory follicles 3.5mm

依靠后裔测定数据和排卵数、产羔数的记录数据, 有报道指出*FecB*基因对排卵数和产羔数性状的影响。

一. 研究背景: *FecB*基因定位

2001年 Wilson 等、Souza 等和 Mulsant 等几乎同时发现, *FecB* 基因所在的绵羊 6号染色体区域同线性的人 4 号染色体 q22 ~ q23 上的**骨形态发生蛋白受体 IB 基因**(bone morphogenetic protein receptor-IB gene, *BMPR-IB*), 与 Booroola Merino 羊的排卵率有关联。

| 野生型 (+/+) | 杂合型 (+/B) | 纯合型 (B/B) |
|-----------|-----------|-----------|
| Q249 | Q249 | Q249R |
| Q249 | Q249R | Q249R |

*BMPR-IB*基因的**A746G**位置的突变,导致249位置处的**谷氨酰胺**转变为**精氨酸**

三. 序列比对：双序列比对 (Needle)

| 物种 | 登录号 | 得分 | 相同位点 | 相似位点 | 空位 |
|------|-------------------|--------|----------------|----------------|-----------|
| 人/绵羊 | O00238/A0A6P3CWH2 | 2654.0 | 494/502(98.4%) | 500/502(99.6%) | 0/502(0%) |

四. BLAST: PBLAST (NCBI)

| | | | | | | | | | |
|---|--|-----------------------------------|------|------|------|--------|--------|-----|--------------------------|
| ✓ | RecName: Full=Bone morphogenetic protein receptor type-1B; Short=BMP type-1B receptor; Short=BMPR-1B; AltN... | Homo sapiens | 1015 | 1015 | 100% | 0.0 | 98.41% | 502 | O00238.1 |
| ✓ | RecName: Full=Bone morphogenetic protein receptor type-1B; Short=BMP type-1B receptor; Short=BMPR-1B; AltN... | Mus musculus | 1014 | 1014 | 100% | 0.0 | 98.61% | 502 | P36898.1 |
| ✓ | RecName: Full=Bone morphogenetic protein receptor type-1B; Short=BMP type-1B receptor; Short=BMPR-1B; AltN... | Gallus gallus | 945 | 945 | 100% | 0.0 | 92.23% | 502 | Q05438.1 |
| ✓ | RecName: Full=Bone morphogenetic protein receptor type-1A; Short=BMP type-1A receptor; Short=BMPR-1A; AltN... | Homo sapiens | 743 | 743 | 97% | 0.0 | 73.73% | 532 | P36894.2 |
| ✓ | RecName: Full=Bone morphogenetic protein receptor type-1A; Short=BMP type-1A receptor; Short=BMPR-1A; AltN... | Mus musculus | 743 | 743 | 97% | 0.0 | 73.93% | 532 | P36895.1 |
| ✓ | RecName: Full=Bone morphogenetic protein receptor type-1A; Short=BMP type-1A receptor; Short=BMPR-1A; AltN... | Rattus norvegicus | 742 | 742 | 100% | 0.0 | 72.67% | 532 | Q78EA7.1 |
| ✓ | RecName: Full=TGF-beta receptor type-1; Short=TGFR-1; AltName: Full=Serine/threonine-protein kinase receptor ... | Rattus norvegicus | 498 | 498 | 95% | 2e-172 | 53.93% | 501 | P80204.1 |
| ✓ | RecName: Full=TGF-beta receptor type-1; Short=TGFR-1; AltName: Full=ESK2; AltName: Full=Transforming growth... | Mus musculus | 495 | 495 | 95% | 2e-171 | 53.91% | 503 | Q64729.1 |
| ✓ | RecName: Full=TGF-beta receptor type-1; Short=TGFR-1; AltName: Full=TGF-beta type I receptor; AltName: Full=T... | Sus scrofa | 490 | 490 | 95% | 3e-169 | 53.39% | 503 | Q5CD18.1 |
| ✓ | RecName: Full=TGF-beta receptor type-1; Short=TGFR-1; AltName: Full=TGF-beta type I receptor; AltName: Full=T... | Bos taurus | 488 | 488 | 96% | 1e-168 | 52.36% | 499 | O46680.1 |
| ✓ | RecName: Full=TGF-beta receptor type-1; Short=TGFR-1; AltName: Full=Activin A receptor type II-like protein kinas... | Homo sapiens | 483 | 483 | 94% | 2e-166 | 53.61% | 503 | P36897.1 |

Family & Domains¹

Domains and Repeats

| Feature key | Position(s) | Description |
|---------------------|-------------|--|
| Domain ¹ | 174 - 203 | GS InterPro annotation |
| Domain ¹ | 204 - 494 | Protein kinase InterPro annotation |

Region

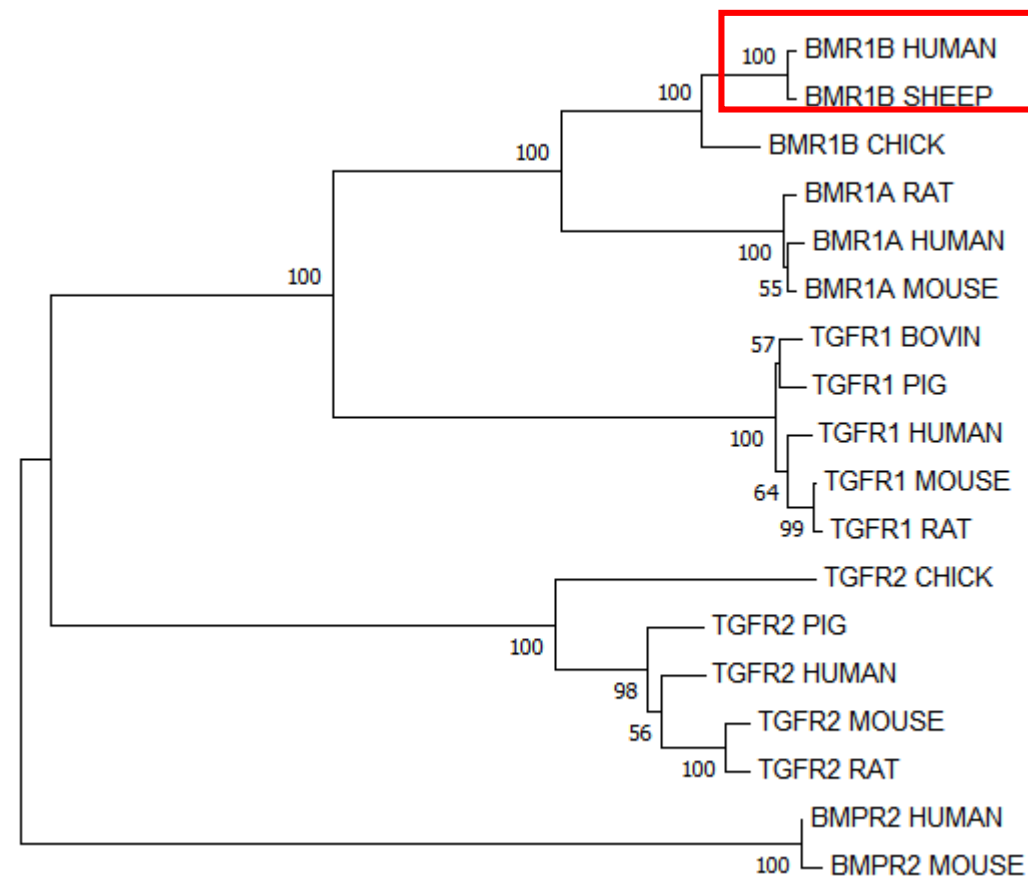
| Feature key | Position(s) | Description |
|---------------------|-------------|--|
| Region ¹ | 1 - 24 | Disordered Sequence analysis |

Sequence similarities²

Belongs to the protein kinase superfamily. TKL Ser/Thr protein kinase family. TGFB receptor subfamily.

五. 系统发育树：不同物种TGF- β 受体家族进化关系（MEGA7）

骨形态发生蛋白受体（BMPRs）家族作为转化生长因子受体（TGF β Rs）超家族中最大的一支，广泛分布在动物体内，依靠卵巢功能性的旁分泌/自分泌调节体系，对哺乳动物**早期胚胎发育**、**卵泡发育**、**生殖激素调节**以及**卵子发生**起重要调节作用。



六. 蛋白结构: Sheep→Human

Structureⁱ

Sheep

No structure information available for A0A6P3CWH2

Human

| | | | | | | | |
|-----------|--------------|-----------|--------|-----|---------|---|-------------------|
| PDB | 3MDY | X-ray | 2.05 Å | A/C | 168-502 | PDBe · RCSB-PDB · PDBj · PDBsum | ↓ |
| AlphaFold | AF-000238-F1 | Predicted | | | 1-502 | AlphaFold | ↓ |



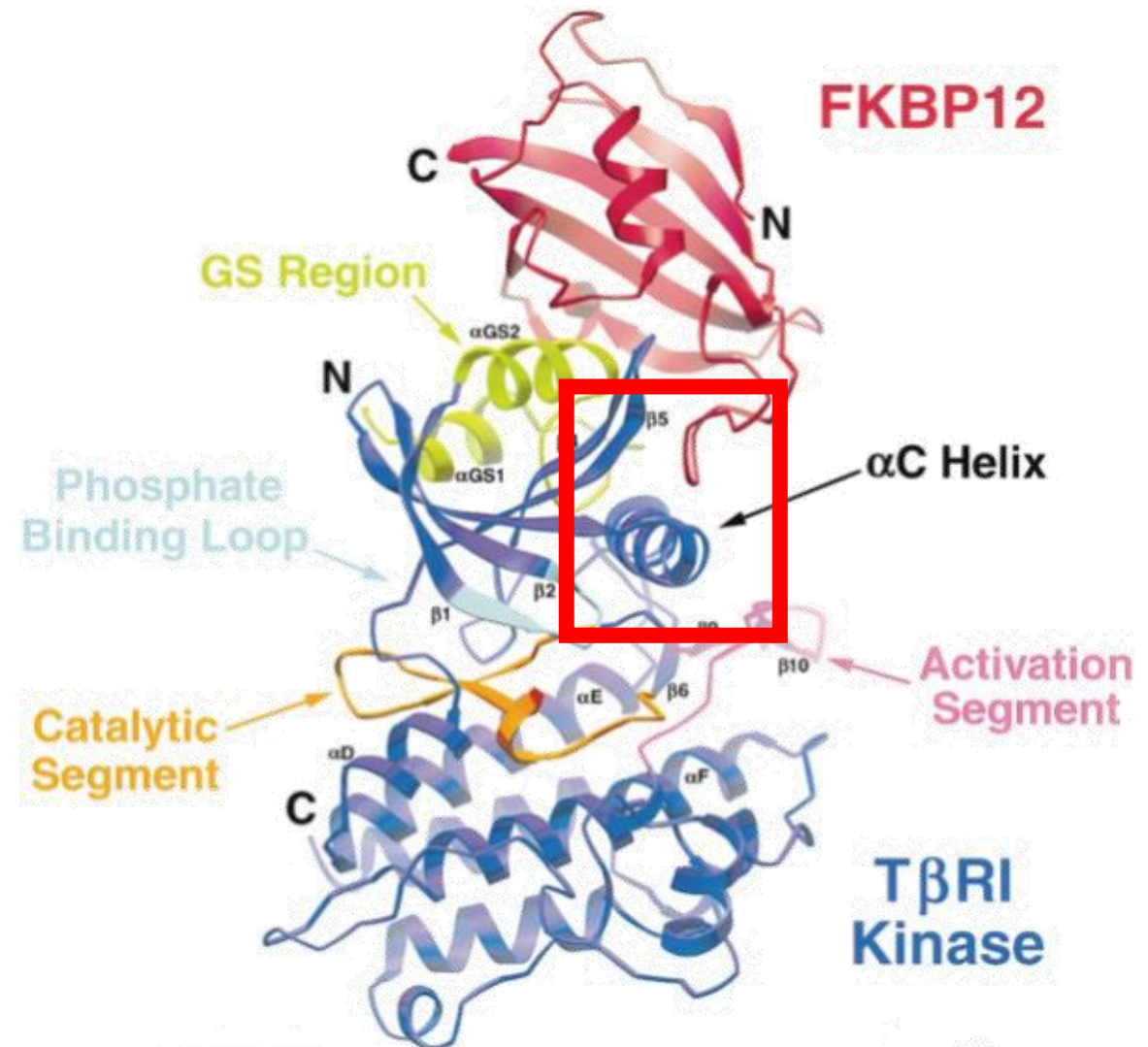
3MDY--PDB数据库

骨形态发生蛋白受体1b (BMPRI1B)与**FKBP12**和LDN-193189复合物细胞质域的晶体结构

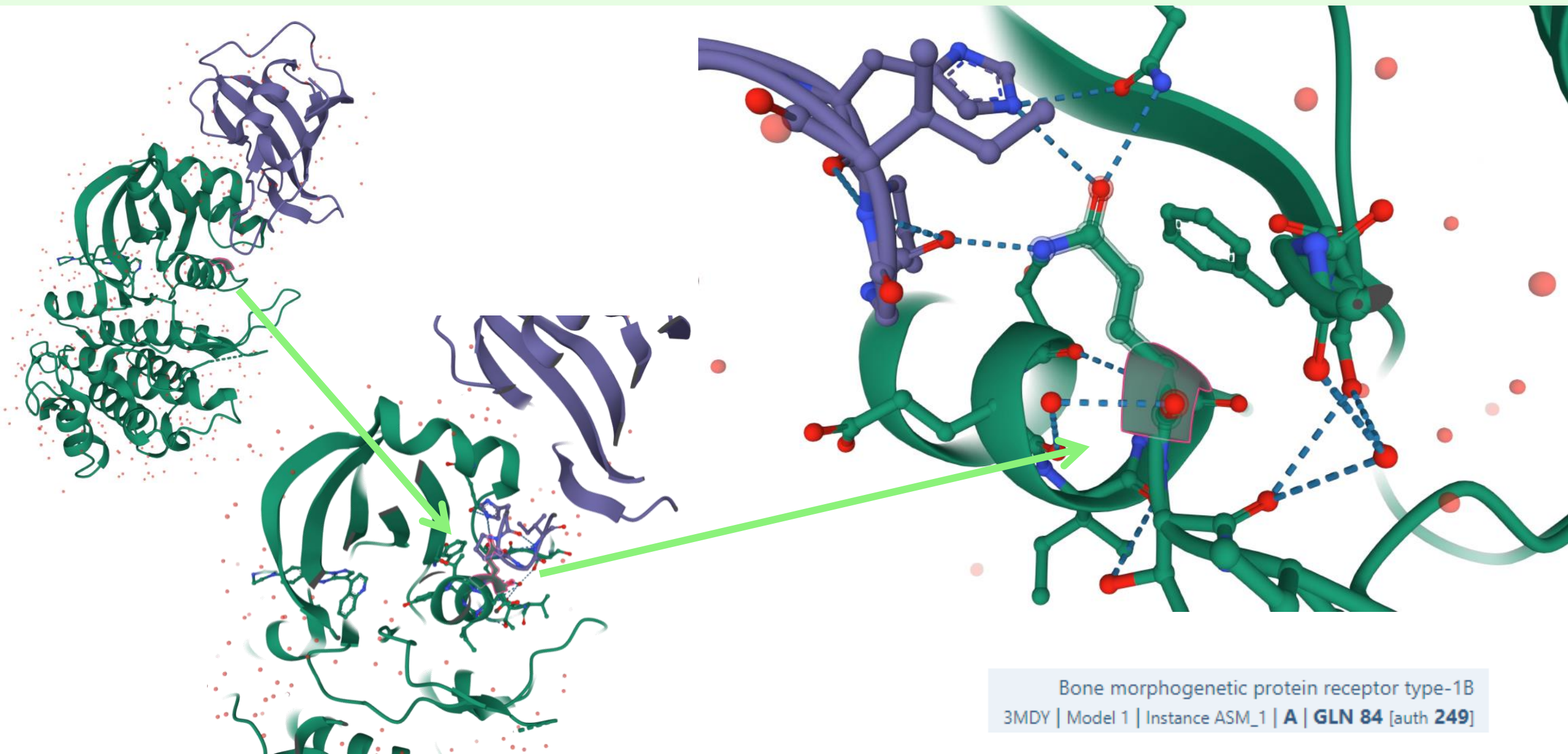
FKBP12是人TGF- β 受体 (T β R-I) 家族成员活性的**反馈调节剂**, 能与BMPRI1B受体**结合**, 抑制BMPRI1B与配体的结合能力。

六. 蛋白结构：Human→Sheep

绵羊BMPRI1B蛋白空间结构根据人TGF- β 受体 (T β R-I) 和克莫司结合蛋白-12 (FKBP12) 构建成为一个**复合模型**。绵羊BMPRI1B的FecB突变 (Q249R) 位于GS结构域和L45环之间, 与人TGF- β 受体结构域中的**Q250残基**相对应, Q250残基位于与FKBP12分子相连的 **α -C螺旋的C端**。



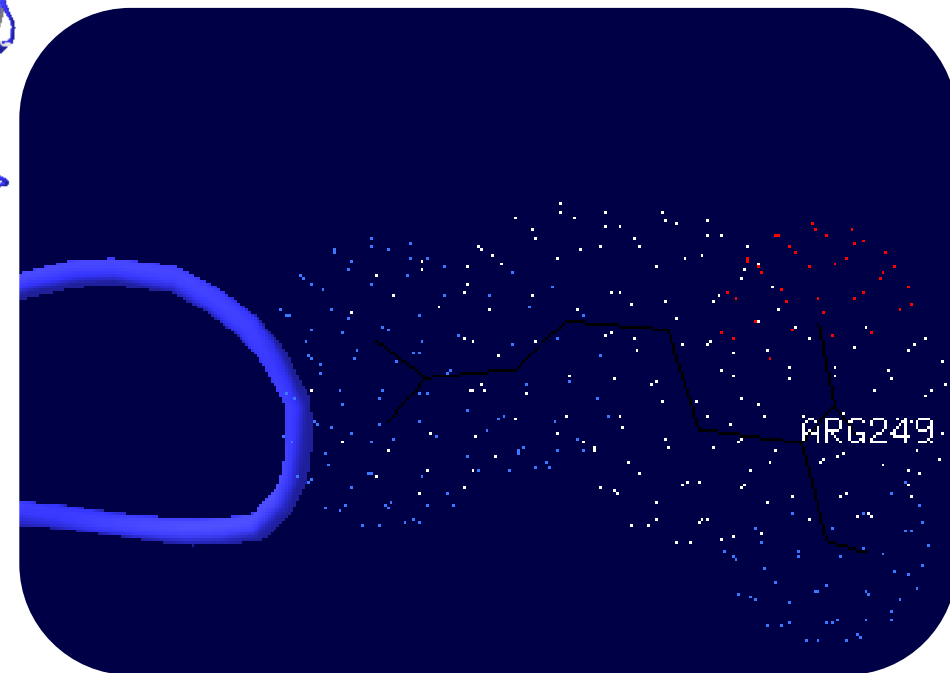
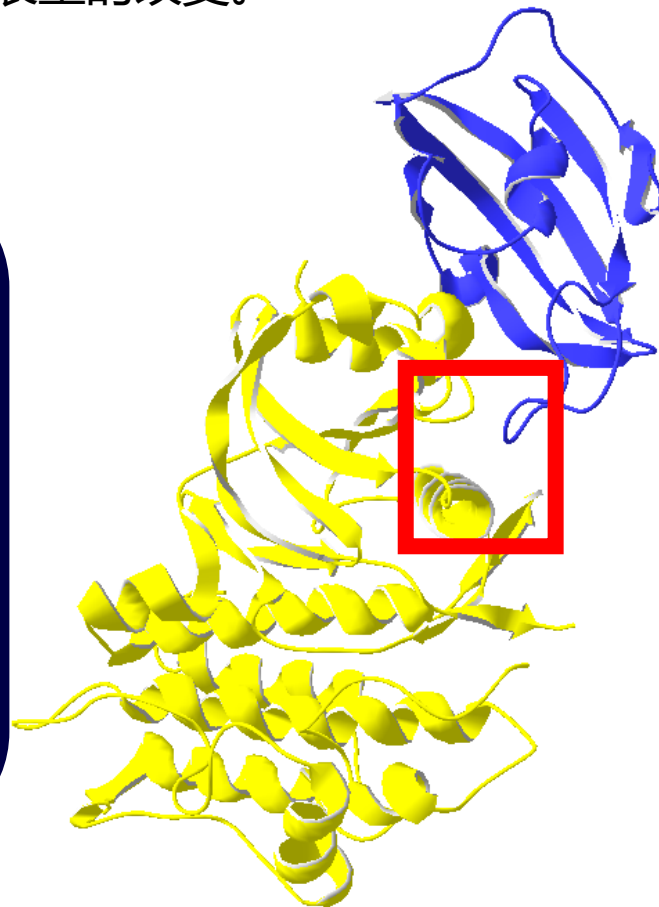
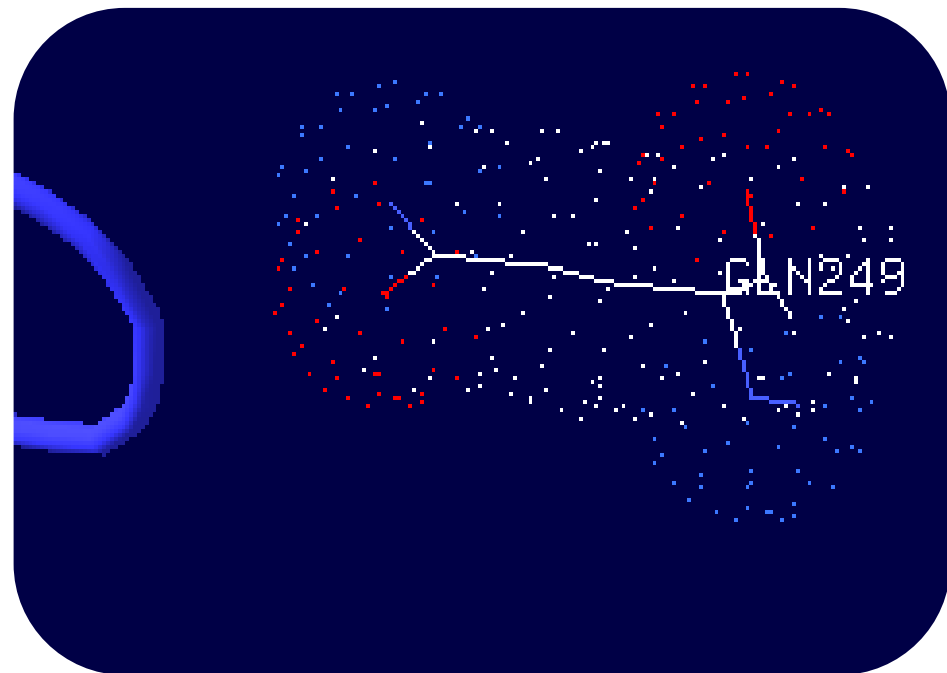
七. 结构改变引起功能改变：PDB (3D-view)



七. 结构改变引起功能改变：PdbViewer

Booroola绵羊的**Q249R**突变使得BMPRII Q250与FKBP12形成氢键，增强了FKBP12与BMPRII之间的 π 电子的相互作用，进而**结合的更紧密**，导致对BMPRII受体活性的**抑制作用增加**。

所以，可以认为Q249R突变使得FKBP12对BMPRII活性**抑制作用的加强**，细胞对BMPRII特异性配体的敏感性变化，最终导致细胞活性的改变，以至于最后表型的改变。



八. 研究展望：技术落地

| 申请号 | 申请日 | 发明名称 | 发明人 |
|----------------|------------|---|---|
| 201510594573.6 | 2015/09/17 | 利用Taqman MGB探针检测绵羊 <i>FecB</i> 基因多态性的方法 | 储明星, 刘秋月, 狄冉, 胡文萍, 王翔宇, 王贵, 潘章源, 赵万民, 李虎山 |
| 201510304092.7 | 2015/06/04 | 利用SNaPshot技术检测绵羊 <i>FecB</i> 基因多态性的方法 | 储明星, 刘秋月, 狄冉, 胡文萍, 王翔宇, 王贵, 潘章源, 赵万民, 李虎山 |



八. 研究展望：机理研究



Transcriptome Analysis Reveals Differentially Expressed Genes and Long Non-coding RNAs Associated With Fecundity in Sheep Hypothalamus With Different FecB Genotypes

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Hypothalamus With Different FecB
Genotypes.

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Small-tailed Han sheep, with different *FecB* genotypes, manifest distinct ovulation rates and fecundities, which are due to differences in reproductive hormones secreted by the hypothalamic–pituitary–ovarian axis. Nevertheless, the function of the hypothalamus against a *FecB* mutant background on increasing ovulation rate is rarely reported. Therefore, we determined the expression profiles of hypothalamus tissue collected from six wild-type (WW) and six *FecB* mutant homozygous (BB) ewes at the follicular and luteal phases by whole-transcriptome sequencing. We identified 53 differentially expressed mRNAs (DEGs) and 40 differentially expressed long non-coding RNAs (DELs) between the two estrus states. Functional annotation analysis revealed that one of the DEGs, PRL, was particularly enriched in the hypothalamic function, hormone-related, and reproductive pathways. The lncRNA–target gene interaction networks and KEGG analysis in combination suggest that the lncRNAs LINC-676 and WNT3-AS cis-acting on *DRD2* and *WNT9B* in different phases may induce gonadotropin-releasing hormone (GnRH) secretion. Furthermore, there were differences of regulatory elements and WNT gene family members involved in the follicular–luteal transition in the reproductive process between wild-type (*WNT7A*) and *FecB* mutant sheep (*WNT9B*). We combined the DEG and DEL data sets screened from different estrus states and genotypes. The overlap of these two sets was identified to select the mRNAs and lncRNAs that have major effects on ovulation. Among the overlapping molecules, seven DEGs and four DELs were involved in the follicular–luteal transition regulated by *FecB* mutation. Functional annotation analysis showed that two DEGs (*FKBP5* and *KITLG*) were enriched in melanogenesis, oxytocin, and GnRH secretion. LINC-219386 and IGF2-

RESEARCH ARTICLE

Hypothalamic Proteome

Proteomics

Proteomics and Systems Biology

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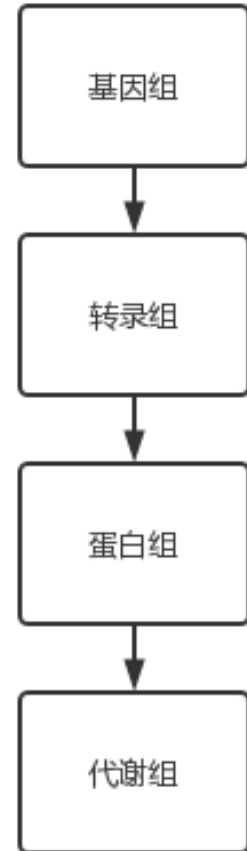
Identification of Prolificacy-Related Differentially Expressed Proteins from Sheep (*Ovis aries*) Hypothalamus by Comparative Proteomics

Zhuangbiao Zhang, Jishun Tang, Ran Di, Qiuyue Liu, Xiangyu Wang, Shangquan Gan,
Xiaosheng Zhang, Jinlong Zhang, Wei Chen, Wenping Hu,* and Mingxing Chu*

Reproduction, as a physiologically complex process, can significantly affect the development of the sheep industry. However, a lack of overall understanding to sheep fecundity has long blocked the progress in sheep breeding and husbandry. In the present study, the aim is to identify differentially expressed proteins (DEPs) from hypothalamus in sheep without *FecB* mutation in two comparison groups: polytocous (PF) versus monotocous (MF) sheep at follicular phase and polytocous (PL) versus monotocous (ML) sheep at luteal phase. Totally 5058 proteins are identified in sheep hypothalamus, where 22 in PF versus MF, and 39 proteins in PL versus ML are differentially expressed, respectively. A functional analysis is then conducted including Gene Ontology and Kyoto Encyclopedia of Genes and Genomes pathway analysis to reveal the potential roles of these DEPs. The proteins ENSOARP00000020097, ENSOARP00000006714, growth hormone (GH), histone deacetylase 4 (HDAC4), and 5'-3' exoribonuclease 2 (XRN2) in PF versus MF, and *bcl-2*-associated athanogene 4 (BAG4), insulin-like growth factor-1 receptor (IGF1R), hydroxysteroid 11-beta dehydrogenase 1 (HSD11B1), and transthyretin (TTR) in PL versus ML appear to modulate reproduction, presumably by influencing the activities of gonadotropin-releasing hormone (GnRH). This study provides an alternative method to identify DEPs associated with sheep prolificacy from the hypothalamus. The mass spectrometry data are available via ProteomeXchange with identifier PXD013822.

1. Introduction

The achievement of reproduction is an essential but complex process involving the central nervous system and endocrine activities, especially those involving hormones released from the hypothalamic–pituitary–gonadal axis.^[1] The hypothalamus, as a critical organ in this axis, initiates the reproductive process through a gonadotropin-releasing hormone (GnRH) signal, which then travels to the pituitary, where follicle-stimulating hormone and luteinizing hormone are induced to function in both the ovary and testis, enabling the synthesis and release of ovarian estradiol and testicular hormone, facilitating folliculogenesis and spermatogenesis, respectively, and maintaining the functions of corpus luteum.^[2–4] Numerous studies involving GnRH and several neuroendocrine factors, such as kisspeptins and neurokinin B, known to be related to GnRH release^[4–6] have been widely studied. Furthermore, some metabolic



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T H A N K S

